

Technical Data Sheet

Innate Immuno-Oncology Protein Panel

Product Information

Material Number:	572615
Size:	2 Tests
Vol. per Test:	NA
Reactivity:	Tested in Development: Human
Storage Buffer:	
Component:	51-9025300
Description:	APC/Myeloid-Cell Protein Panel
Size:	1 Test (2 ea)
Vol. per Test:	NA
Component:	51-9025299
Description:	NK-Cell Protein Panel
Size:	1 Test (2 ea)
Vol. per Test:	NA
Component:	51-9025129
Description:	Tumor Protein Panel
Size:	1 Test (2 ea)

Description

The BD® OMICS-One Innate Immuno-Oncology Protein Panel consists of 6 single tubes, 2 tubes of Tumor Protein Panel (1 test/tube) containing 30 different specificities against major Tumor markers, 2 tubes of NK-Cell Protein Panel (1 test/tube) containing 30 different specificities against major NK-Cell markers, and 2 tubes of APC/Myeloid-Cell Protein Panel (1 test/tube) containing 30 different specificities against major APC/Myeloid-Cell markers. Designed and optimized to work on the BD Rhapsody™ System, the Innate Immuno-Oncology Protein Panel is tested to work seamlessly alongside the BD Rhapsody™ Whole Transcriptome Analysis (WTA) Assay, Targeted mRNA Assay, BD® Single-Cell Multiplexing Kit (SMK), BD® Intracellular CITE-seq (IC-AbSeq) Assay, and BD Rhapsody™ TCR/BCR Next Multiomic Assay for human. The individual antibodies were each conjugated to an oligonucleotide that contains a specific antibody barcode sequence flanked by a polyA tail on the 3' end and a common PCR handle (PCR primer binding site) on the 5' end. All AbSeq barcode sequences were generated in-silico with minimal sequence similarity to the human genomes, have low predicted secondary structure, and have high Hamming distance within the BD antibody-oligo portfolio, to allow for sequencing error correction and unique mapping. The polyA tail of the oligonucleotide allows the barcode sequence to be captured by BD Rhapsody™ Enhanced Cell Capture Beads. The 5' PCR handle allows for efficient library generation for various sequencing platforms. Each individual antibody exists at an optimal concentration within the 84-plex panel to enable superior target and population resolution.

The Innate Immuno-Oncology Protein Panel is designed with SMART technology. SMART technology helps lower sequencing cost while increasing data resolution by attenuating antibodies that target high-expressing primary markers and allowing reallocation of sequencing reads to markers expressed at lower levels. With SMART technology, markers low in expression can be quantified without having to do deeper sequencing and incurring high sequencing cost. There are six specificities attenuated in the Innate Immuno-Oncology Protein Panel, CD2 and CD31, CD44, CD45, HLA-DR, and HLA-ABC.

Preparation and Storage

Store at 2-8°C and protected from prolonged exposure to light.

Do not freeze.

Application Notes

Application

Single Cell 3' Sequencing	Qualified
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Recommended Assay Procedure:

BD Biosciences

bdbiosciences.com

United States 877.232.8995 Canada 866.979.9408 Europe 32.2.400.98.95 Japan 0120.8555.90 Asia Pacific 65.6861.0633 Latin America/Caribbean 55.11.5185.9995

For country contact information, visit bdbiosciences.com/contact

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572615 Rev.



This reagent is provided lyophilized in a pre-titrated format. Each test can stain up to 2 million cells.

1. Remove one tube of BD® OMICS-One Tumor Protein Panel, one tube of BD® OMICS-One NK-Cell Protein Panel and one tube of BD® OMICS-One APC/Myeloid-Cell Protein Panel from foil bags and bring up to room temperature for 5 minutes.
2. Make sure all pellets are located at the bottom of the tubes. If not, briefly centrifuge to collect the contents at the tube bottom.
3. For each tube, add 35 µL of nuclease-free water to the bottom of the tube and allow antibodies to reconstitute for 5 minutes at room temperature.
4. Place the reconstituted antibodies on ice until the cells are ready for staining.
Note: Reconstitute antibody **immediately** before cell staining. Prolonged incubation of reconstituted antibody may increase the non-specific background.
5. For BD® AbSeq Ab-Oligo drop-in of 25 plex or lower, prepare the BD® AbSeq labeling MasterMix in 1.5-mL LoBind tube on ice.
Note: For drop-in with more than 25 plex, reach out to technical support for calculation.

If sequential labeling with Sample Tags or no Sample Tags, prepare BD® AbSeq labeling MasterMix for drop-ins as follows:

Component	1 sample (µL)	1 sample + 30% overage (µL)	2 samples + 30% overage (µL)
Per BD® AbSeq Ab-Oligo	2.0	2.6	5.2
Total of BD® AbSeq Ab-Oligo	2.0 × N*	2.6 × N	5.2 × N
FBS† (catalog number 554656)	70 - (2.0 × N)	91 - (2.6 × N)	182 - (5.2 × N)
Total	70	91	182

If co-labeling with Sample Tags, prepare BD® AbSeq labeling MasterMix for drop-ins as specified as follows:

Component	1 sample (µL)	1 sample + 30% overage (µL)	2 samples + 30% overage (µL)
Per BD® AbSeq Ab-Oligo	2.0	2.6	5.2
Total of BD® AbSeq Ab-Oligo	2.0 × N*	2.6 × N	5.2 × N
FBS† (catalog number 554656)	50 - (2.0 × N)	65 - (2.6 × N)	130 - (5.2 × N)
Total	50	65	130

* N = number of drop-in antibodies. N = 0 if there are no drop-in antibodies.

† FBS = BD Pharmingen™ Stain Buffer.

6. Pipet-mix the BD® AbSeq labeling MasterMix for drop-ins. Briefly centrifuge to collect the contents at the bottom, and place back on ice.
7. For sequential labeling with Sample Tags or no Sample Tags, for each sample, combine the three tubes containing 35 µL reconstituted NK-Cell Protein Panel solution, 35 µL reconstituted APC/Myeloid-Cell Protein Panel solution, and 35 µL reconstituted Tumor Protein Panel solution. Then add 70 µL BD® AbSeq labeling MasterMix of drop-ins to the tube containing 105 µL reconstituted Tumor, NK-Cell and APC/Myeloid-Cell Protein Panels solution to make a total volume of 175 µL.
For co-labeling with Sample Tags, for each sample, combine the three tubes containing 35 µL reconstituted NK-Cell Protein Panel solution, 35 µL reconstituted APC/Myeloid-Cell Protein Panel solution, and 35 µL reconstituted Tumor Protein Panel solution. Then add 50 µL BD® AbSeq labeling MasterMix of drop-ins and 20 µL Sample Tag to the tube containing total 105µL Tumor, NK-Cell and APC/Myeloid-Cell Protein Panels solution to make a total volume of 175 µL.
8. Pipet-mix the mixture, briefly centrifuge to collect the contents at the tube bottom, and place back on ice.
9. Centrifuge cells at 400 × g for 5 minutes. If Fc Block is used, proceed to step 10. If Fc Block is not used, skip to step 11.
10. (Optional) For samples containing myeloid and B lymphocytes, BD Biosciences recommends blocking nonspecific Fc Receptor-mediated false-positive signals with Human BD Fc Block (catalog number 564220).
 - a. To perform blocking, pipet the Fc Block MasterMix into a new 1.5-mL LoBind tube on ice:

Component	1 sample (µL)*	1 sample + 20% overage (µL)
FBS† (catalog number 554656)	20.0	24.0
Fc Block‡ (catalog number 564220)	5.0	6.0
Total	25.0	30.0

* Sufficient for up to 1 million cells. To block more cells, adjust the volume.

† FBS = BD Pharmingen™ Stain Buffer.

‡ Fc Block = BD Pharmingen™ Human BD Fc Block.

- b. Pipet-mix the Fc Block MasterMix and briefly centrifuge. Place on ice.
 - c. Remove the supernatant from the cells without disturbing the pellet.
 - d. Resuspend the cells in 25 μ L of Fc Block MasterMix.
 - e. Incubate the cells at room temperature (15°C to 25°C) for 10 minutes.
 - f. Add 175 μ L of BD® AbSeq labeling MasterMix from Step 8 into the cell suspension. Pipet-mix and proceed to Step 12.
11. Remove the supernatant from the cells without disturbing the pellet. Add 25 μ L Stain Buffer (FBS) to the 175 μ L of BD® AbSeq labeling MasterMix from Step 8 to make a total volume of 200 μ L. Resuspend the cell pellet in 200 μ L total volume. Pipet-mix.
 12. Transfer the cells with BD® AbSeq labeling MasterMix into a new 5-mL polystyrene Falcon tube.
 13. Stain the cells on ice for 30 minutes.
 14. Add 3-4 mL Stain Buffer (FBS) to labelled cells and pipet-mix.
 15. Centrifuge at 400 \times g for 5 minutes.
 16. Uncap the tube and invert to decant supernatant into biohazardous waste. Keep the tube inverted and gently blot on a lint-free wiper to remove residual supernatant from tube rim.
 17. Repeat steps 14–16 twice more for a total of three washes.
 18. Resuspend the final washed cell pellet in 620 μ L cold Sample Buffer from the BD Rhapsody™ Enhanced Cartridge Reagent V3 (catalog number 667052) and proceed to single cell capture with on-cartridge washing described in substeps a–c. Refer to the *BD Rhapsody™ HT Single-Cell Analysis System Single-Cell Capture and cDNA Synthesis Protocol* (Doc ID 23-24252) or *BD Rhapsody™ HT Xpress System Single-Cell Capture and cDNA Synthesis Protocol* (Doc ID 23-24253) for additional details.
- Note:** Perform on-cartridge washing after cell settling (8-minute incubation) as described in the following sub-steps.
- a. At the protocol section of “Loading cells in BD Rhapsody™ 8-Lane Cartridge”, after cell load, incubate the cartridge in the dark at room temperature for 8 minutes.
 - b. Place the cartridge on the BD Rhapsody™ HT Xpress and perform the On-Cartridge Wash steps as follows:

Material to load	Volume (μ L) 1 lane	Pipette Mode
Air	380	Prime/Wash
Cold Sample Buffer	380	Prime/Wash
Air	380	Prime/Wash
Cold Sample Buffer	380	Prime/Wash

- c. (Optional) Perform the scanner step: Cell Load Scan, if using *BD Rhapsody™ HT Single-Cell Analysis System Single-Cell Capture and cDNA Synthesis Protocol* (Doc ID 23-24252). No need for 8-minute delay before scanning.

Warning: All biological specimens and materials are considered biohazardous. Handle as if capable of transmitting infection and dispose using proper precautions in accordance with federal, state, and local regulations. Never pipette by mouth. Wear suitable protective clothing, eyewear, and gloves.

List of all 30 Human AbSeq specificities included in the BD® OMICS-One Tumor panel:

Specificity	Clone	Oligo ID	BD® AbSeq Barcode Sequence
CD274 (PD-L1) [†]	MIH1	AHS0004	ATCGTAAGGCTCGTGGTTCGTAAGTAAGTTCGTATC
CD279 (PD-1)	EH12.1	AHS0014	ATGGTAGTATCACGACGTAGTAGGGTAATTGGCAGT
CD45 [†]	HI30	AHS0040	GTGCGAAATGGCGGAATGTTATCTGCGAATGTAGTC
CD324 (E-cad)	67A4	AHS0041	GATATGAATGGGTTGCGGTTAAAGTCGTAATGGTT
CD24	ML5	AHS0042	ACTTTGGGTTGAGCGCATGATTATTCGTGACACTTT
CD90	5E10	AHS0045	GACTATATGTACGGTGAATTCGGGATCCTGCGCT
CD34	581	AHS0061	TGGGTGATTACGGTTAGTTTATGCGCGAAGGTGTT
CD117	YB5.B8	AHS0064	GGATTAGTTGTCGTTATAGGGAGTGCCTTCTTAGCG
HLA-A, B, C	G46-2.6	AHS0066	GATATGCATGGCGAGTAGGTAGAACGAAGCTTAGGT
CD54*	HA58	AHS0076	AAGAGAATATATGCGTGCCTGTTGTTAAGGGAATGCGT
CD29	MAR4	AHS0080	TGGTAAGGTGGTTGCGAGTAAGTAGCGGTGAGTTGT
CD47	B6H12	AHS0087	TGTTAGGTTTCGACGATTTATGTGTAGATCCGCAAGG
CD326 (EpCam)	EBA-1	AHS0089	TTGAGCGTAAAGTTGCGTCCGGTAATTGAGTTGCGT
CD66	B1.1/CD66	AHS0094	GTCTGCGCAAGGTAAGCTAAGTAACGAAAGGATCT
CD133	W6B3C1	AHS0103	TTTGGTATTGGCACGGTTTGTAGCGAGTTGACGGTC
CD26	M-A261	AHS0109	TGTAGGTTGCGCGGTTATTAGGGTATTATCGATCTG
CD155	TX24	AHS0111	GCGGTGGATCGATGGGTATAGTTGTAATTTGCGTC
CD146	P1H12	AHS0127	AGGTTATTTAGGTGACGGTTGATTGACGAGAGAGG
C-MET	3D6	AHS0132	AGCGTGAGTTGTCGGTAGTTAATTATCGGAGAGTTT
ITGRN BTA 7	F1B504	AHS0158	TTTCAGTTTGGTCGCAGTTAAGGTATCGATGGGTC
CD44	L178	AHS0167	GTGATTGATTAGGACAGTTTCGTTGCTTAGTAGTGGG

CD31 (PECAM1)*	WM59	AHS0170	CTAAGGGACGTAATTGAGTTTCGGTGATCGCAGTTT
EphB2	2H9	AHS0176	TATTGCGGGTAGGATTGTCTCGAAGCGTAGGTAGC
Vista†	MIH65.RMAB	AHS0187	ATCAGGGAATCTCGGTAAGTTAAACGTGTATAGTGC
PDPLN	LPMAB-17	AHS0192	TTTATGAGTATTACGTCTGTTGCGATTGTTGGCGGT
NOTCH1	MHN1-519	AHS0214	CGTAGTAGGAGCGTGTTCATCGGCATTATCGTTTG
CD325 (n-Cad)	8C11	AHS0223	TAGGATGAGTTTCGTAAGTAAGGTAGTCGTATGGCT
CD58	1C3	AHS0237	TTGGTGAGTATTGGTGCAGTATGCGGGATGTTTG
EGFR	EGFR.1	AHS0241	ATATGATTGATGCGGGTTAGCCTACAGATTGAGTT
CD227 (MUC1)	HMF62	AHS0247	AGTGCATGTTTAGTAGGTGTGAGTCGTTAGATATTC

List of all 30 Human AbSeq specificities included in the BD® OMICS-One NK-Cell panel:

Specificity	Clone	Oligo ID	BD® AbSeq Barcode Sequence
CD11b‡	M1/70	AHS0005	ATCGTTATTGTTGTAGTTTCGCCGGTTTGAGTAGT
CD56	NCAM16.2	AHS0019	AGAGGTTGAGTCGTAATAATAATCGGAAGCGTTGG
CD38	HIT2	AHS0022	GTC AACGATGGGTAGCGGTAGAAAACGGAAC TGG
CD27	M-271	AHS0025	TGTCGGGTTTAGCGAATTGGGTTGAGTCACGTAGGT
CD2	RPA-2.10	AHS0029	AAACGTAGATTAGAGCCGGGTATGTCGCAACTGATT
CD16‡	3G8	AHS0053	TAAATCTAATCGCGGTAACATAACGGTGGGTAAGGT
CD184 (CXCR4)	12G5	AHS0060	CAGTGTTTAGAGCGGGTTCATATGTCGTTTAGAGG
CD49d	9F10	AHS0063	TAGGGTGACTTAGCGATTGATGCGTATGTTGGGCG
CD314 (NKG2D)	10D11	AHS0065	TTGAAATGCGATGAGACGTAGAGCGCATGAGGTAGC
CD335 (NKP46)	9E2/NKP46	AHS0068	CAATTTGTTGCGGTTTAGTAGTCGTCGTTTATGGG
CD54*	HA58	AHS0076	AAGAGAATATATGCGTGCGTTGTTAAGGGAATGCGT
CD226	DX11	AHS0079	GAGTTTATGATTGCTTTCTTCGGTAGTTCGTCGCTT
CD94	HP-3D9	AHS0085	GAGGTTAGGATAGGTGTACGGGTCGAGTTGAATCT
CD336 (NKP44)	p44-8	AHS0090	AATGCAAACGATATCACGAAGGGTAGTACACGACGG
CD49a	SR84	AHS0101	ATGACACGAATGCGACGAGAGGCGAAATAGGTTGGT
CX3CR1	2A9-1	AHS0125	GGGTTACGAGGTTTAAAGCGGTAGTATAGGATGCC
CD122	Mik-β3	AHS0146	TTAAAGAGATTTCGTGGGTATTGGCGCAGTCATTCCT
CD140b (PDGFR)	28D4	AHS0151	GACAACATTTAGGACGTGACGAGAGATATAGCTTC
CD248	B1/35	AHS0156	ATCACTTATTTGTTTGGAGGGTTCGTAGGCGTTGC
CD63	H5C6	AHS0157	TGCAGCGTTAGGACCAAGCGTTTACCCTAGAATATT
CD140a	α-R1	AHS0160	TTACTGACTTTTCGGACGTTGGTTACTTAGGGTTATG
CD31 (PECAM1)*	WM59	AHS0170	CTAAGGGACGTAATTGAGTTTCGGTGATCGCAGTTT
CD96	6F9	AHS0194	CTAATGTAAGAGCGGACGTTTGGGCACTATATGTTT
CD161 (KLRB1)	HP-3G10	AHS0205	TTTATGACGATTAGTTGTGCGGCATAGGAGGTGTTT
CD158b (KIR)	DX27	AHS0209	CGTAGGAGGATTTTCGTCGATGGGTTTGTAGCGTTC
CD158e1	DX9	AHS0211	AGGTTTATTGCGGCATTAGGCGTCATATAGTAGGTG
CD337/Nkp30	P30-15	AHS0213	GGTAACTGACATGACGGAGCGATAATTTCTGGCGGT
CD3	UCHT1	AHS0231	AGCTAGGTGTTATCGGCAAGTTGTACGGTGAAGTCG
CD329 (Siglec-9)	E10-286	AHS0239	CGGGCGGAAGATAGGATAATAGGTAACGTCAAATG
CD106	51-10C9	AHS0251	TCTGATTTAGCGGGTGGACGTATTATAGTGATTGGC

List of all 30 Human AbSeq specificities included in the BD® OMICS-One APC/Myeloid-Cell panel:

Specificity	Clone	Oligo ID	BD® AbSeq Barcode Sequence
CD103	BER-ACT8	AHS0001	AAATAGTATCGAGCGTAGTTAAGTTGCGTAGCCGTT
CD274 (PD-L1)†	MIH1	AHS0004	ATCGTAAGGCTCGTGGTTCGTAAGTAAGTTCGTATC
CD11b‡	M1/70	AHS0005	ATCGTTATTCGTTGATGTTCCGCCGTTTGAGTAGT
CD123	7G3	AHS0020	ACAGTTTAGTAGGACGTGAGGTATCGCGAGAATGCC
HLA-DR	G46-6	AHS0035	TGTTGGTTATTCGTTAGTGCATCCGTTTGGGCGTGG
CD14	MPHIP9	AHS0037	TGGCCCCGTGAGCGCAATGTGAGATCGTAATAAGT
CD45†	HI30	AHS0040	GTGCGAAATGGCGGAATGTTATCTCGGAATGTAGTC
CD33	WM53	AHS0044	GTGTTAGTGATTTGATAGGACGCGTTACGAGAGATT
CD80	L307.4	AHS0046	GAGGGTAACGGGTGTCCAAATATCGGCTGTGTAAGT
CD16‡	3G8	AHS0053	TAAATCTAATCGCGTAACATAACGGTGGGTAAGGT
CD64	10.1	AHS0055	TTGTGCGCGCTAGTATGGTTATCTCGAGTGAAAGTC
CD11c	B-LY6	AHS0056	ATGCGTTGCGAGAGATATGCGTAGGTTGCTGATTGG
CD163	GHI/61	AHS0062	TATTATGTGCGAACTATGGTATCCGTATTGAGGGCT
CD195 (CCR5)	2D7/CCR5	AHS0070	ATGGTTTAGTCGTACGTGGGTTTAGATTGGCGGTGC
CD206	19.2	AHS0072	GCTGGTTATCGTTTGAGAGTCGGTATGGAATGCGGT
CD32	FLI8.26	AHS0073	GGTTGTAGGTGCGGAATATAAGCGTCGTTGAGGTGT
CD273	MIH18	AHS0075	TGAGTAACCGTATGTAATCCGTAATCGTAGAAGCGC
CD141	1A4	AHS0083	TGGAAGTAAGTATGGGTCGGCGTAAATTGTGCGTGT
CD1c	F10/21A3	AHS0088	ATAGATTACATTCTGTTTACGCTTGGGTTCCGTCCTG
CD40	5C3	AHS0117	GGTGAATTGGGCTAGAACGTATATGCGGTAAGGCG
FCeR1a	AER-37	AHS0129	GATATGGCGTGATGGTAGGTTCCGTTTAAGTTAGCG
CD169	7-239	AHS0133	CATTAAGCACGAAGGGTATAGGTAGGAACGGTTGGC
CD36	IVC7	AHS0135	AATTGTAGTAGTCCGGTGTATGTAGAGTAGGCGTTT
CD115 (CSF1R)	9-4D2-1E4	AHS0136	CTGGTGGCGGCGAATTTGGTTACGACATATAGGGTT
CD162	KPL-1	AHS0139	CCAGATAGGCGATAGTGTTAGGAGCGATTAGTGTG
CD85K	ZM3.8	AHS0179	AGTAGTCGTAGTTGGCGTGAATTGGGCTTATATCTG
VISTA†	MIH65.RMAB	AHS0187	ATCAGGGAATCTCGGTAAGTTAAACGTGTATAGTGC
CD15	W6D3	AHS0196	ATAGGCATGGACGACGTAGATAATAAGTGCCGGGTT
CD192 (CCR2)	LS132.1D9	AHS0208	CATGAGTGAGGCGATATAGTGAGCGGTTTGTAGATT
CD116	Hgmcsfr1-M1	AHS0238	CTTAGTTGTAGGATCGAGAGTAGGTTGTCATTGCGT

* Tumor and NK-Cell Protein Panels contain the same CD54 and CD31 (PECAM1) antibody.

† Tumor and APC/Myeloid-Cell Protein Panels contain the same CD274 (PD-L1), CD45 and VISTA antibody.

‡ NK-Cell and APC/Myeloid-Cell Protein Panels contain the same CD11b and CD16 antibody.

Suggested Companion Products

<u>Catalog Number</u>	<u>Name</u>	<u>Size</u>	<u>Clone</u>
554656	Stain Buffer (FBS)	500 mL	(none)
564220	Human BD Fc Block™	0.25 mg	Fc1
633801	Whole Transcriptome Analysis (WTA) Amplification Kit	1 Each	(none)
633774	Targeted mRNA and AbSeq Amplification Kit	1 Each	(none)
667058	TCR/BCR Next Amplification Kit	1 Each	(none)
633773	cDNA Kit	1 Each	(none)
666262	8-Lane Cartridge	1 Each	(none)
667052	Enhanced Cartridge Reagent Kit V3	1 Each	(none)
633781	Hu Single Cell Sample Multiplexing Kit	1 Each	(none)
633849	Flex Single-Cell Multiplexing Kit A, Flex Sample Tag 1-6	1 Each	(none)
633850	Flex Single-Cell Multiplexing Kit B, Flex Sample Tag 7-12	1 Each	(none)
633851	Flex Single-Cell Multiplexing Kit C, Flex Sample Tag 13-18	1 Each	(none)
633852	Flex Single-Cell Multiplexing Kit D, Flex Sample Tag 19-24	1 Each	(none)
666625	BD® Rhapsody™ HT Xpress Package		(none)
633701	BD Rhapsody Scanner		(none)
570742	Intracellular AbSeq Buffer Kit	1 Each	(none)
570911	OMICs-Guard Sample Preservation Buffer	50 mL	(none)
570750	AbSeq Enhancer Kit	1 Each	(none)
570751	RNase Inhibitor	1 Each	(none)

Product Notices

1. This reagent is provided lyophilized in a pre-titrated format. Go to <https://www.bdbiosciences.com/en-us/resources/protocols/single-cell-multiomics> for additional BD Rhapsody™ protocols.
2. The production process underwent stringent testing and validation to assure that it generates a high-quality conjugate with consistent performance and specific binding activity. However, verification testing has not been performed on all conjugate lots.
3. Go to <https://abseq-ref-gen.genomics.bd.com/> to access AbSeq reference files in FASTA format for bioinformatics analyses.
4. Caution: Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing. Follow state and local guidelines when disposing of hazardous waste.
5. For U.S. patents that may apply, see bd.com/patents.
6. Read and understand the safety data sheets (SDSs) before handling chemicals. To obtain SDSs, go to regdocs.bd.com or contact BD Biosciences technical support at scomix@bd.com.