

 **BD Rhapsody™ HT Single-Cell
Analysis System**
Extended-Lysis Single-Cell Capture and
cDNA Synthesis
Protocol

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History

Revision	Date	Change made
23-24984(01)	2025-09	Initial release.
23-24984(02)	2026-03	Added cartridge step and illustrations.
23-24984(03)	2026-09	Updated with corrections for customer-impacted issues.

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Introduction

This protocol describes cell loading in the BD Rhapsody™ 8-Lane Cartridge and single-cell capture with the BD Rhapsody™ HT Single-Cell Analysis System.

For complete instrument procedures and safety information, see the *BD Rhapsody™ HT Single-Cell Analysis System Instrument User Guide*.

Required and recommended materials

For a complete list of materials, see the instrument user guide.

Symbols

The following symbols are used in this guide:

Symbol	Description
	Important information for maintaining measurement accuracy or data integrity.
	Noteworthy information.
	Procedural stopping point.
	Biological hazard.

Required reagents

Material	Supplier	Catalog no.
BD Rhapsody™ Enhanced Cartridge Reagent Kit v3	BD Biosciences	667052
BD Rhapsody™ 8-Lane Cartridge	BD Biosciences	666262
BD Rhapsody™ cDNA Kit	BD Biosciences	633773
BD® OMICS-One Dual Index Kit ^a	BD Biosciences	571899
Absolute ethyl alcohol, molecular-biology grade	Major supplier	–
Nuclease-free water	Major supplier	–
BD Pharmingen™ Calcein AM ^b	BD Biosciences	564061
BD Pharmingen™ DRAQ7™ ^b	BD Biosciences	564904
Dimethyl sulfoxide (DMSO)	Major supplier	–
70% ethyl alcohol or 70% isopropyl alcohol ^c	Major supplier	–
a. Additional indexing primers for high-throughput library preparation workflows require the BD® OMICS-One Dual Index Kit. b. Protect Calcein AM and DRAQ7™ from light. Avoid multiple freeze-thaw cycles of Calcein AM. See manufacturer's storage recommendations. c. To clean the BD Rhapsody™ HT Xpress System and the BD Rhapsody™ Scanner, see the <i>BD Rhapsody™ Single-Cell Analysis System Installation and Maintenance Guide</i>. Instead of 70% alcohol, 10% (v/v) bleach can be used.		

Recommended consumables

Material	Supplier	Catalog no.
Gilson™ PIPETMAN™ DIAMOND Tipack™ Filter Tips, 100-1200 µL for BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette) (Recommended) Or, ZAP™ SLIK 1000 µL Low Retention Aerosol Filter Pipet Tips for BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette) (Alternative)	Thermo Fisher Scientific Labcon	F171803 or F171803G 1177-965-008-9
60 mL reagent reservoir self-standing ^a	BD Biosciences	666626
Reagent reservoir (sterile, non-pyrogenic, RNase/DNase free), 10 mL	VistaLab	3054-1012 3054-1013
Reagent reservoir (sterile, non-pyrogenic, RNase/DNase free), 25 mL	VistaLab	3054-1002 3054-1003
Falcon® tube with cell strainer cap	Corning	352235
Corning® 96-well polypropylene Cluster Tube, 8-tube strip format, sterile ^b	Corning	4413
DNA LoBind® tubes, 1.5-mL	Eppendorf	30108051
DNA LoBind® tubes, 2.0-mL	Eppendorf	022431048
Low-retention, filtered pipette tips (20 µL, 200 µL, 1000 µL)	Major supplier	–
INCYTO™ disposable hemocytometer	INCYTO	DHC-N01-5
Deep 96-Well 2 mL Polypropylene plate	Major supplier	–
Pre-moistened cleaning wipes with 70% ethyl alcohol or 70% isopropyl alcohol.	Major supplier	–
Lint-free wipes	Major supplier	–
a. Waste collection container for the BD Rhapsody™ HT Xpress System. b. These are the bead retrieval tubes to be used with the BD Rhapsody™ HT Xpress System.		

Required equipment

Material	Supplier	Catalog no.
BD Rhapsody™ HT Xpress Package ^a	BD Biosciences	666730
BD Rhapsody™ Scanner ^b	BD Biosciences	633701
Hemocytometer adapter ^c	BD Biosciences	633703
BD Rhapsody™ P8xP1200µL Pipette - HTX ^d	BD Biosciences	500066280
BD Rhapsody™ P1200µL Pipette - HTX ^{ce}	BD Biosciences	500066148
Microcentrifuge for 1.5–2.0-mL tubes	Major supplier	–
Centrifuge and rotor with adapters for 5-mL Falcon® tubes and 15-mL tubes.	Major supplier	–
Eppendorf ThermoMixer® C ^f	Eppendorf	5382000023
SmartBlock™ Thermoblock 1.5-mL ^f	Eppendorf	5360000038
Incubator at 37 °C	Major supplier	–
Pipettes (P10, P20, P200, P1000)	Major supplier	–
Multi-channel pipette (P200)	Major supplier	–
Vortexer	Major supplier	–
Digital timer	Major supplier	–
6-Tube magnetic separation rack for 1.5-mL tubes Or, 12-Tube magnetic separation rack Or, Invitrogen™ DynaMag™-2 magnet	New England Biolabs New England Biolabs Thermo Fisher Scientific	S1506S S1509S 12321D
a. Includes BD Rhapsody™ P8xP1200µL Pipette - HTX and Hamilton 60-mL Waste Reservoirs. b. Includes a Hemocytometer adapter. c. Included with the BD Rhapsody™ Scanner. Item can be ordered separately. d. Part of the BD Rhapsody™ HT Single-Cell Analysis Package. Items can be ordered separately. e. Only required if not using the BD Rhapsody™ P8xP1200µL Pipette - HTX. f. For cDNA synthesis after cartridge cell capture.		

Preparation list

Item		BD Part Number	Preparation and Handling	Storage
Equilibrate to room temperature				
Cartridge Wash Buffer 1		650000060	Equilibrate to room temperature.	4 °C
Cartridge Wash Buffer 2		650000061		
	RT Buffer	650000067	Equilibrate to room temperature 30 minutes before setting up reverse transcription (RT). Centrifuge briefly. Keep on ice until ready.	–20 °C
	dNTP	650000077		
	RT 0.1 M DTT	650000068		
	Nuclease-Free Water	650000076		
	10X Exonuclease I Buffer	650000071		
	Bead Resuspension Buffer	650000066		
Place on ice				
BD Rhapsody™ Enhanced Cell Capture Beads		91-1294	Keep on ice until ready.	4 °C
Sample Buffer		650000062		
Lysis Buffer		650000064		
Bead Wash Buffer		650000065		
1.0 M DTT		650000063		
Leave at –25 °C to –15 °C				
	Bead RT/PCR Enhancer	91-1082	Centrifuge briefly before adding to mix.	–20 °C
	RNase Inhibitor	650000078		
	Reverse Transcriptase	700026321		
	Exonuclease I	650000072		
Obtain				

Item	BD Part Number	Preparation and Handling	Storage
BD Rhapsody™ 8-lane Cartridge	666262	—	Ambient
2 mM Calcein AM		Thaw and protect from light.	–20 °C
0.3 mM DRAQ7™		Protect from light.	4 °C
100% ethyl alcohol			
Ice bucket			
1.5 mL DNA LoBind® tubes			
INCYTO™ disposable hemocytometer			
Cell suspension			
Set up			
Thermomixer at 80 °C			
Thermomixer at 37 °C			

Best practices

- Always use low-retention filtered pipette tips and LoBind tubes.
- Perform single-cell capture and cDNA synthesis in a pre-amplification workspace.
- Prepare cells as close to cell loading as possible.
- Change pipette tips before every pipetting step.
- Keep reagents on ice unless instructed otherwise.

Guidelines for good pipetting

The following guidelines apply to the BD Rhapsody™ P8xP1200µL Pipette and BD Rhapsody™ P1200µL Pipette:

- BD Rhapsody™ P8xP1200µL Pipette: Push the tip holder into the tips while rocking back and forth to ensure a firm and airtight seal.
- BD Rhapsody™ P1200µL Pipette: Push the tip holder into the tip using a slight twisting motion to ensure a firm and airtight seal.
- Hold the pipette vertically and immerse the tip 2-4 mm in the liquid. Make sure that the reservoir has enough volume to avoid aspirating air.
- Press the push button to aspirate the set volume of liquid. Wait for 2-3 seconds. Then withdraw the pipette tip from the liquid. Ensure that every tip aspirates the same volume of the liquid before loading in the cartridge.
- While removing the pipette from the reservoir, draw the tip along the inside surface of the vessel.
- When dispensing, ensure that the pipette tips are seated perpendicular to the inlet of the BD Rhapsody™ 8-Lane Cartridge.
- Press the push button to dispense. While tips are still inserted in the cartridge inlet, wait for at least a few seconds before releasing the push button to expel any residual liquid from the tip.

Before you begin

- Visually inspect the Lysis Buffer for any precipitation. If precipitation is present, incubate the Lysis Buffer at room temperature (15–25 °C) for 1 hour. Invert to mix but do not vortex. Once the solution is clear, place the Lysis Buffer on ice.
- Open the DTT tube while holding the tube vertically. The solution is overlain with an inert/non-oxygen-containing gas, and a non-vertical tube will allow the inert gas to pour off. If not loading 4 or 8 lanes at the same time, after opening the DTT tube once, seal and store the tube at –20 °C or aliquot for single use in 0.5-mL microcentrifuge tubes and store at –20 °C.
- Thaw Calcein AM and once at room temperature (15–25 °C), resuspend Calcein AM in 503.0 µL dimethyl sulfoxide (DMSO) for a final stock concentration of 2 mM. Follow the manufacturer's instructions and protect from light.
- After opening the cartridge packet just before the start of the experiment, ensure that you prepare only the lanes you intend to use in the cartridge.
- If cell preparation takes 4 hours or longer, begin preparing cells before cartridge preparation.
- For guidance on preparing a single-cell suspension, see *Preparing Single-Cell Suspensions Protocol*.

- If your biological sample contains red blood cell contamination, red blood cell lysis is required. See *Preparing Single-Cell Suspensions Protocol*.

Priming and treating BD Rhapsody™ 8-Lane Cartridge

Prime and treat the BD Rhapsody™ 8-Lane Cartridge. For detailed instructions, see the instrument user guide.



- Ethyl alcohol priming of the cartridge followed by air purge provides full coverage of the array during the Prime/Wash step (step 2 on the following table).
- Random bubbles (<3 mm diameter in size) that occur during the Priming steps do not affect cartridge performance.
- In lanes where bubbles >3 mm in size are observed, aspirate and dispense air using the Prime/Wash mode and repeat step 1 with 100% ethyl alcohol. Only do this in the Priming steps.
- Uneven fluidic front observed on different lanes does not affect cartridge performance.
- Discard used tips even if residual volume is observed after dispensing.



It is recommended to use a P20 pipette to aspirate buffer pooling at the inlet. Aspirate at an angle to avoid accidental aspiration of buffer volume in the microwell array.

- 1 Aliquot 100% ethyl alcohol and cartridge reagent buffers in 10-mL or 25-mL reagent reservoirs as shown in the following table, depending on the number of lanes used. Do not aliquot for single lane.

Component	For 1 lane (mL)	For 2 lanes (mL)	For 3 lanes (mL)	For 4 lanes (mL)	For 5 lanes (mL)	For 6 lanes (mL)	For 7 lanes (mL)	For 8 lanes (mL)
100% ethyl alcohol	0.05	2.00	2.00	2.00	2.00	2.00	2.00	2.00

Component	For 1 lane (mL)	For 2 lanes (mL)	For 3 lanes (mL)	For 4 lanes (mL)	For 5 lanes (mL)	For 6 lanes (mL)	For 7 lanes (mL)	For 8 lanes (mL)
Cartridge Wash Buffer 1	0.76	3.50	5.25	7.00	8.75	10.50	12.25	14.00

Component	For 1 lane (mL)	For 2 lanes (mL)	For 3 lanes (mL)	For 4 lanes (mL)	For 5 lanes (mL)	For 6 lanes (mL)	For 7 lanes (mL)	For 8 lanes (mL)
Cartridge Wash Buffer 2	0.38	2.00	3.00	4.00	5.00	6.00	7.00	8.00

- 2 Remove cartridge from foil pouch. Retain foil pouch and dessicant to store partially used cartridge.
- 3 Place waste collection container and cluster tube in the **BD Rhapsody™ HT Xpress System**.
- 4 Carefully peel off the seals on the cartridge inlet depending on the number of lanes to be used.
- 5 See the following table for instrument slider position.

BD Rhapsody™ HT Xpress System slider	Position
Front slider	Waste
Retrieval slider	Inactive (Back)

Step no.	Material to load	Volume (μL) 1 lane	Pipette Mode	Incubation at room temperature
1	100% ethyl alcohol	50	EtOH Prime	—
2	Air	380	Prime/Wash	—
3	Room temp. Cartridge Wash Buffer 1	380	Prime/Wash	1 min
4	Air	380	Prime/Wash	—
5	Room temp. Cartridge Wash Buffer 1	380	Prime/Wash	3 min
6	Air	380	Prime/Wash	—
7	Room temp. Cartridge Wash Buffer 2	380	Prime/Wash	≤4 hr

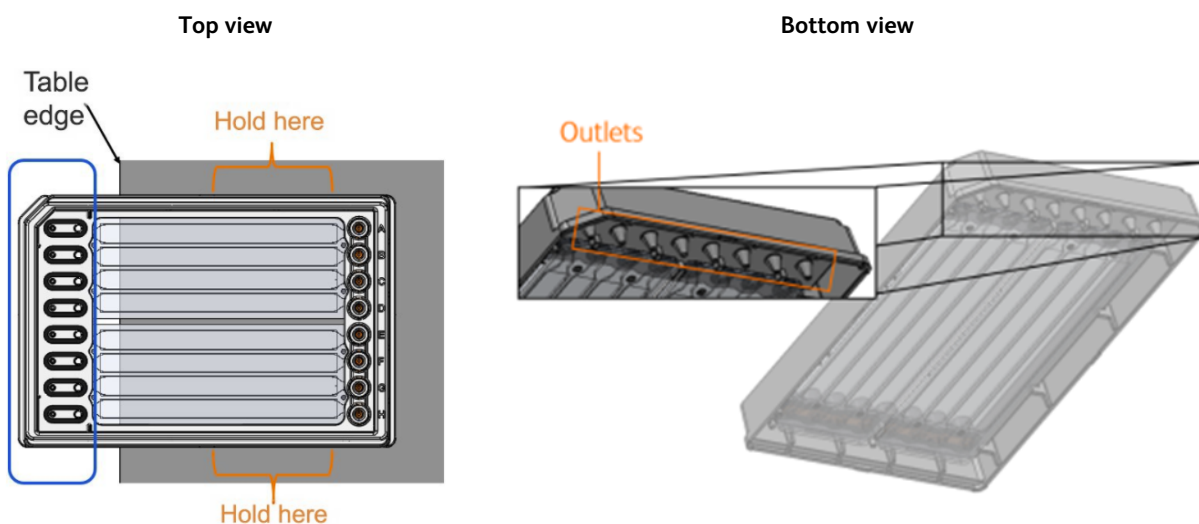
6



Before the cell loading step (after airing out the Cartridge Wash Buffer 2), remove the cartridge from the HT Xpress System.

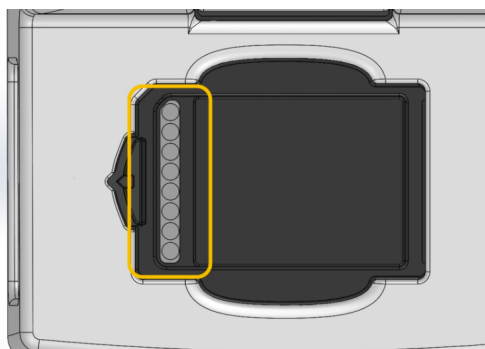
Keep the cartridge level. Do not invert or tilt the cartridge. Place it on a lab bench with the outlet area hanging out as shown in the following top view illustration. Use a lint-free wipe to blot away any excess liquid from around the outlets. Do not blot directly on the outlets or for an extended period of time, because this might wick buffer from the lane (indicated in the following bottom view illustration).

Liquid creeping might occur during priming steps due to low surface tension reagents used in priming steps. It will not affect cartridge performance. However, it is important to remove the residues of the priming buffers from the outlet area.





In addition, check the BD Rhapsody™ HT Xpress station. If there is any wetness or liquid droplets spotted on the station, use a lint-free wipe to dry the area, especially near the outlet area highlighted in the yellow rectangle in the following illustration. Verify that the station remains dry during the workflow to prevent liquid creeping.



Staining cells with viability markers



Protect Calcein AM and DRAQ7™ from light until ready to use.

- 1 If cells are not resuspended in cold Sample Buffer, centrifuge the cell suspension at 400g for 5 minutes, aspirate the supernatant, and leave ~20 µL of residual supernatant. Add up to 620 µL total volume of cold Sample Buffer.



Performance might be impacted if samples are not in Sample Buffer. For rare samples that are not resuspended in Sample Buffer before cell loading, proceed at your own risk, or contact technical support.

- 2 Add 3.1 µL of 2 mM Calcein AM and 3.1 µL of 0.3 mM DRAQ7™ to 620 µL of cell suspension (1:200 dilution) in cold Sample Buffer.
- 3 Gently pipet-mix.
- 4 Incubate at 37 °C in the dark for 5 minutes.
- 5 Filter cells through Falcon® Tube with cell strainer cap.



For low-abundance or low-volume samples, filtering is optional at this step.

- 6 Determine the desired number of cells to capture in the BD Rhapsody™ 8-Lane Cartridge. See the *BD Rhapsody™ HT Single-Cell Analysis Instrument User Guide* for a table containing estimated multiplet rates based on the number of captured cells on retrieved BD Rhapsody™ Enhanced Cell Capture Beads.
- 7 Count cells immediately using the scanner.
 - a Ensure cells are well suspended by gently pipet-mixing.
 - b Pipet 10 µL into the INCYTO™ disposable hemocytometer.

Keep the remaining cells on ice, and protect from light.

Counting and preparing single-cell suspension for cartridge loading

For detailed instructions on counting cells with the BD Rhapsody™ Scanner, see the instrument user guide.

- 1 Insert the hemocytometer into the Hemocytometer Adapter, and tap **Scan**.
- 2 Place the adapter on the scanner tray, and tap **Continue**.
- 3 Select **Hemocytometer** for the protocol, and select or enter the experiment name, sample name, and user.
- 4 Tap **Side A** or **Side B**, then **Start Side A Scan** or **Start Side B Scan (Cell Count)**.
- 5 After the scan is complete, tap **OK**.
- 6 Tap **Scan**, and enter a new sample name to scan the other side of the hemocytometer. Repeat steps 4–5, or tap **Eject** and remove the adapter. Tap **Return Home**.
- 7 Tap **Analysis** and experiment name to view the total cell concentration and cell viability.
- 8 Proceed as follows:
 - If the cell concentration is $\leq 1,000$ cells/ μ L, proceed to step 9.
 - If the cell concentration is $> 1,000$ cells/ μ L, dilute the cell suspension in cold Sample Buffer to ~ 200 – 800 cells/ μ L. Repeat steps 1–7, and then step 9.
- 9 Tap **Prepare** to display the Samples Calculator screen.
- 10 Dispose of the hemocytometer.



Minimize the time between cell pooling and single-cell capture.

- 11 Use the Samples Calculator to obtain stock cell and buffer volumes from the scanner to prepare a cell suspension of 380 μ L.
- 12 Select the correct Cartridge Type. For the BD Rhapsody™ 8-Lane Cartridge, use 0120.
- 13 Prepare the cell suspension according to the displayed volumes on the scanner.

Ensure the stock solution of each sample is well suspended by gently pipet-mixing before pooling.
- 14 Transfer each tube of the prepared single-cell suspension to a 96-deep well plate for multiple lane loading. Keep on ice.

Minimize the time between cell pooling and single-cell capture.

Loading cells in BD Rhapsody™ 8-Lane Cartridge

- 1 Load the cartridge with materials listed using the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette):

Material to load	Volume (µL) 1 lane	Pipette mode
Air	380	Prime/Wash
• Gently pipet-mix with a multi-channel pipette to completely resuspend the cells • Set the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette) to Load mode. • Immediately load.		
Cell suspension	320	Load



Air bubbles that might appear at the inlet or outlet of the cartridge (outside of the microwell array) do not affect cartridge performance.

- 2 If necessary, wipe condensation from the top cartridge surface for optimal scanning.
- 3 Incubate at room temperature (15–25 °C) for 8 minutes. To incubate the cartridge on the scanner, enter a time delay of 8 minutes before tapping **Start Cell Load Scan** (Cell Load step).
- 4 During the 8-minute incubation, prepare the BD Rhapsody™ Enhanced Cell Capture Beads.
See [Preparing BD Rhapsody™ Enhanced Cell Capture Beads](#) in the following section.
- 5 Image the cells in the cartridge. Perform the scanner step: **Cell Load**. For more information, see the instrument user guide.
- 6 After the scan is complete, tap **OK**, then **Eject**. Remove the cartridge from the scanner. After the scan, confirm the analysis is running.



Optional: If using AbSeq, Sample Tag, or ATAC-Seq workflow, it is recommended to do two washes after the 8 minute incubation, according to the following table.

Material to load	Volume (µL) 1 lane	Pipette mode
Air	380	Prime/Wash
Cold Sample Buffer	380	Prime/Wash
Air	380	Prime/Wash
Cold Sample Buffer	380	Prime/Wash

Preparing BD Rhapsody™ Enhanced Cell Capture Beads



Do not vortex samples containing BD Rhapsody™ Enhanced Cell Capture Beads. Gently mix suspensions with the beads by pipette only.

Keep BD Rhapsody™ Enhanced Cell Capture Beads on ice before use.



Use low-retention pipette tips and LoBind tubes. Keep the beads cold, and pipet-mix only.

- 1 Place the beads tube on the magnet for 1 minute.
- 2 Remove and discard the storage buffer.
- 3 Remove the tube from the magnet, and pipet 380 μ L of cold Sample Buffer into the tube.
- 4 Pipet-mix, and place on ice.
- 5 Transfer each tube of the beads to a 96-deep well plate for multiple lane loading. Keep on ice until Cell Load scan is complete.
- 6 After the Cell Load scan and analysis is running, proceed to [Loading and washing BD Rhapsody™ Enhanced Cell Capture Beads](#) in the following section.

Loading and washing BD Rhapsody™ Enhanced Cell Capture Beads

- 1 Place the cartridge on the BD Rhapsody™ HT Xpress System.
- 2 Load the cartridge with the materials listed using the BD Rhapsody™ P8xP1200 μ L Pipette (or BD Rhapsody™ P1200 μ L Pipette):

Material to load	Volume (μ L) 1 lane	Pipette mode
Air	380	Prime/Wash
• Set the BD Rhapsody™ P8xP1200μL Pipette (or BD Rhapsody™ P1200μL Pipette) to Mix mode, gently mix cell capture beads by pressing the push button six times (waiting for aspiration or dispense to complete each time) or until the beads are completely resuspended. Make sure that the pipette tips are reaching the bottom of the wells to mitigate the chance of introducing air bubbles. Discard used pipette tips. • With a new set of pipette tips, set the pipette to Load mode. • Immediately load. Check the pipette tips to make sure that there are no air bubbles inside the tips before loading. Otherwise, dispense in the 96-deep well plate and aspirate with a new set of pipette tips.		
BD Rhapsody™ Enhanced Cell Capture Beads	320	Load

- 3 Incubate the cartridge at room temperature (15–25 °C) for 3 minutes.
- 4 Perform the scanner step: **Bead Agitation**.

- 5 After Bead Agitation is complete, tap **OK**, then **Eject**.
- 6 Remove the cartridge from the scanner.
- 7 Place the cartridge on the BD Rhapsody™ HT Xpress System.
- 8 Aliquot Sample Buffer in 10 mL reagent reservoir as shown in the following table, depending on the number of lanes used. Do not aliquot for single lane.

Component	For 1 lane (mL)	For 2 lanes (mL)	For 3 lanes (mL)	For 4 lanes (mL)	For 5 lanes (mL)	For 6 lanes (mL)	For 7 lanes (mL)	For 8 lanes (mL)
Sample Buffer	0.38	2.00	2.80	3.60	4.30	5.10	5.90	6.60

- 8 Set the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette) to **Prime/Wash** mode.
- 9 Load each lane used in the cartridge with the materials listed using the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette).

Material to load	Volume (µL) 1 lane	Pipette mode
Air	380	Prime/Wash
Cold Sample Buffer	380	Prime/Wash
Air	380	Prime/Wash
Cold Sample Buffer	380	Prime/Wash

- 10 Perform the scanner step: **Bead Wash**.
- 11 After the scan is complete, tap **OK**, then **Eject**.
- 12 Remove the cartridge from the scanner.
- 13 After the scan, confirm the analysis is running.

Lysing cells



Avoid bubbles.

If all 4 or 8 lanes are not loaded at the same time, seal the opened DTT tube and store at -20°C , or aliquot into single-use 0.5-mL microcentrifuge tubes and store at -20°C .

- 1 Add 75.0 μL of 1 M DTT to one 15-mL Lysis Buffer bottle. Check Add DTT box.



Use the Lysis Buffer with DTT within 24 hours, and then discard.

- 2 Briefly vortex the lysis mix, and aliquot in the 10-mL or 25-mL reagent reservoir as shown in the following table, depending on the number of lanes used. Do not aliquot for single lane. Place on ice.

Component	For 1 lane (mL)	For 2 lanes (mL)	For 3 lanes (mL)	For 4 lanes (mL)	For 5 lanes (mL)	For 6 lanes (mL)	For 7 lanes (mL)	For 8 lanes (mL)
Lysis Buffer	0.28 / 1.0	3.75	5.60	7.50	9.40	11.25	13.10	15.00



The volumes shown in the table are the required volumes for the cell lysis and bead retrieval steps.

- 3 Place the cartridge on the BD Rhapsody™ HT Xpress System.
- 4 Set the BD Rhapsody™ P8xP1200 μL Pipette (or BD Rhapsody™ P1200 μL Pipette) to **Lysis** mode.
- 5 Load the cartridge with the materials listed using the BD Rhapsody™ P8xP1200 μL Pipette (or BD Rhapsody™ P1200 μL Pipette):

Material to load	Volume (μL)	Pipette mode
Lysis Buffer with DTT	280	Lysis

- 6 Incubate at room temperature ($15\text{--}25^{\circ}\text{C}$) for 10 minutes.



Maintain the recommended lysis time for best performance.



Before retrieval, remove extra buffer that has pooled at the inlet with a P20 pipette to minimize overflow. Aspirate at an angle to avoid accidental aspiration of buffer volume in the microwell array.

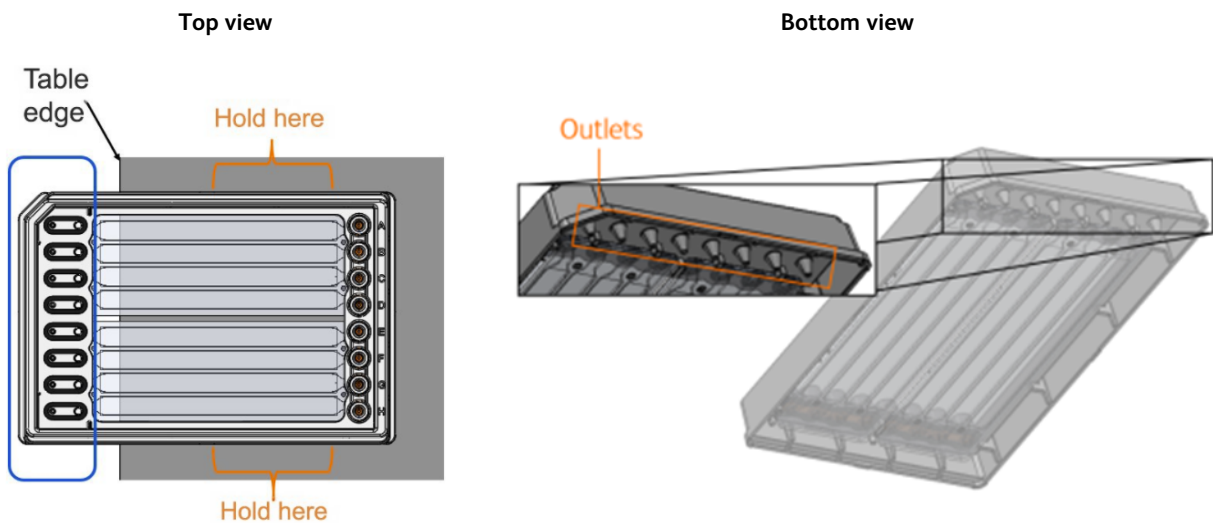
Retrieving BD Rhapsody™ Enhanced Cell Capture Beads

Before Retrieval, remove the cartridge from the HT Xpress System.

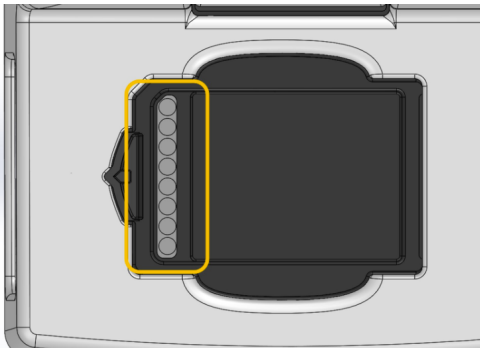


Keep the cartridge level. Do not invert or tilt the cartridge. Place it on a lab bench with the outlet area hanging out as shown in the following top view illustration. Use a lint-free wipe to blot away any excess liquid from around the outlets. Do not blot directly on the outlets or for an extended period of time, because this might wick buffer from the lane (indicated in the following bottom view illustration).

It is important to wipe off the residues from the outlet area before the Retrieval step. Neglecting this step might result in liquid creeping that could lead to sample loss.



In addition, check the BD Rhapsody™ HT Xpress station. If there is any wetness or liquid droplets spotted on the station, use a lint-free wipe to dry the area, especially near the outlet area highlighted in the yellow rectangle in the following illustration. Verify that the station remains dry during the workflow to prevent liquid creeping.



- 1 Place the cluster tube, 8-tube strip format in the BD Rhapsody™ HT Xpress System drawer. Label the tubes appropriately.
- 2 Move the front slider to **BEADS** on the BD Rhapsody™ HT Xpress System.
- 3 Gently pull the top RETRIEVAL slider toward and on top of the BD Rhapsody™ 8-Lane Cartridge (ACTIVE). Make sure that the Retrieval magnet is in contact with the BD Rhapsody™ 8-Lane Cartridge.
- 4 Leave the Retrieval magnet in the ACTIVE position for 1 minute.
- 5 Set the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette) to **Retrieval** mode.
- 6 Aspirate 1,000 µL of Lysis Buffer with DTT using the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette).
- 7 Press down on the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette) to seal against the gasket.
- 8 Push back the top RETRIEVAL magnet (INACTIVE), and immediately load 1,000 µL of Lysis Buffer with DTT.
- 9 Remove the pipette from the gasket, and discard the tip.
- 10 Move the front slider to **OPEN**, and remove the cluster tube with the bottom adapter to a flat, secure surface.



Handle cluster tubes with care. The cluster tubes contain BD Rhapsody™ Enhanced Cell Capture Beads with hybridized bioproducts.

- 11 Move the front slider to **WASTE**.
- 12 Blot the outlet drip on the bottom of the cartridge with a lint-free wipe to remove residual liquid. Perform the scanner step: Retrieval.
- 13 After the scan is complete, tap **OK**, then **Eject**. Remove the cartridge from the scanner and tap **Return Home**. After the scan, confirm the analysis is running.
- 14 Immediately proceed to [Washing BD Rhapsody™ Enhanced Cell Capture Beads](#) in the following section.
- 15 Keep the partially used cartridge on a flat surface while performing Washing BD Rhapsody™ Enhanced Cell Capture Beads and Performing Reverse Transcription.
- 16 Perform [Washing used lanes and BD Rhapsody™ 8-Lane Cartridge storage procedure](#) on [page 26](#) during reverse transcription incubation.



All surfaces that come in contact with biological specimens can transmit potentially fatal disease. Use universal precautions when cleaning surfaces. Wear suitable protective clothing, eyewear, and gloves.

Washing BD Rhapsody™ Enhanced Cell Capture Beads

- 1 Remove the cluster tube from the bottom adapter. Gently pipet-mix the beads and transfer them to a new 1.5-mL LoBind tube. Keep on ice.
- 2 If there are still beads left in the cluster tube, rinse the cluster tube with 100 µL of Lysis Buffer with DTT, and transfer to the 1.5-mL LoBind tube from the previous step.
- 3 Place the tube on the 1.5-mL magnet for ≤ 2 minutes. Remove the supernatant.



Avoid leaving Lysis Buffer or bubbles in the tube. Lysis Buffer may cause the reverse transcription reaction to fail.

- 4 Remove the tube from the magnet, and pipet 1.0 mL of cold Bead Wash Buffer into the tube. Pipet-mix.



When multiple samples are being processed, it is advised to keep the tubes on ice when they are not being washed.

- 5 Place the tube on the 1.5-mL magnet for ≤ 2 minutes. Remove the supernatant.
- 6 Remove the tube from the magnet, and pipet 1.0 mL of cold Bead Wash Buffer into the tube. Pipet-mix, and place on ice.



Start reverse transcription ≤ 30 minutes after washing the retrieved BD Rhapsody™ Enhanced Cell Capture Beads with Bead Wash Buffer.



CRITICAL! If performing the TCR and/or BCR assay, stop here and proceed with respective protocol to continue cDNA synthesis.

Performing reverse transcription

- 1 Ensure that the SmartBlock™ Thermoblock for ThermoMixer® C is at 37 °C.
- 2 In the pre-amplification workspace, pipet the following reagents into a new 1.5-mL or 2.0-mL LoBind tube on ice:

cDNA mix

Cap	Component	For 1 lane (μL)	For 1 lane + 20% overage (μL)	For 4 lanes + 20% overage (μL)	For 8 lanes + 20% overage (μL)
	RT Buffer	40.0	48.0	192.0	384.0
	dNTP	20.0	24.0	96.0	192.0
	RT 0.1 M DTT	10.0	12.0	48.0	96.0
	Bead RT/PCR Enhancer	12.0	14.4	57.6	115.2
	RNase Inhibitor	10.0	12.0	48.0	96.0
	Reverse Transcriptase	10.0	12.0	48.0	96.0
	Nuclease-Free Water	98.0	117.6	470.4	940.8
	Total	200.0	240.0	960.0	1,920.0

- 3 Gently vortex mix, briefly centrifuge, and place back on ice.
- 4 Place the tube of washed BD Rhapsody™ Enhanced Cell Capture Beads on the 1.5-mL tube magnet for ≤2 minutes.
- 5 Remove and discard the supernatant.
- 6 Remove the tube from the magnet and pipet 200 μL of cDNA mix into the beads. Pipet-mix.
Keep the prepared cDNA mix with beads on ice until the suspension is transferred in the next step.
- 7 Transfer the bead suspension to a new 1.5-mL LoBind tube.
- 8 Incubate the bead suspension on the thermomixer at 1,200 rpm and 37 °C for 20 minutes.



Shaking is critical for this incubation.

- 9 During reverse transcription incubation, view the image analysis to see if the analysis metrics passed.
- 10 Place the tube on ice.

Treating sample with Exonuclease I

- 1 Set one thermomixer to 37 °C and a second thermomixer to 80 °C.



Exonuclease I inactivation temperatures above 80 °C can result in the loss of AbSeq molecules, thus a heat block should not be used for this step. If only one thermomixer is available, allow it to equilibrate to 80 °C before starting the inactivation incubation.

- 2 In the pre-amplification workspace, pipet the following reagents into a new 1.5-mL or 2.0-mL LoBind tube on ice:

Exonuclease I mix

Cap	Component	For 1 lane (μL)	For 1 lane + 20% overage (μL)	For 4 lanes + 20% overage (μL)	For 8 lanes + 20% overage (μL)
	10X Exonuclease I Buffer	20.0	24.0	96.0	192.0
	Exonuclease I	10.0	12.0	48.0	96.0
	Nuclease-Free Water	170.0	204.0	816.0	1,632.0
	Total	200.0	240.0	960.0	1,920.0

- 3 Gently vortex mix, briefly centrifuge, and place back on ice.
- 4 Place the tube of BD Rhapsody™ Enhanced Cell Capture Beads with cDNA mix on the 1.5-mL tube magnet for ≤2 minutes. Remove and discard the supernatant.
- 5 Remove the tube from the magnet, and pipet 200 μL of Exonuclease I mix into the tube. Pipet-mix.
- 6 Incubate the bead suspension on the thermomixer at 1,200 rpm and 37 °C for 30 minutes.



If only one thermomixer is available, allow it to equilibrate to 80 °C before starting the inactivation incubation. Place the samples on ice until that temperature is reached.

- 7 Incubate the bead suspension on the thermomixer (no shaking) at 80 °C for 20 minutes.



Do not exceed this inactivation temperature and incubation time.

- 8 Place the tube on ice for ~1 minute. Briefly centrifuge.
- 9 Place the tube on the magnet for ≤1 minute until clear. Remove and discard the supernatant.
- 10 Remove the tube from the magnet, and pipet 200 μL of cold Bead Resuspension Buffer into the tube.

11 Pipet-mix.

Exonuclease I-treated beads can be stored at 2–8 °C for up to 1 year.

12 Proceed to library preparation.

Washing used lanes and BD Rhapsody™ 8-Lane Cartridge storage procedure

- 1 Ensure that the front slider has moved to **WASTE** on the BD Rhapsody™ HT Xpress System.
- 2 Aliquot nuclease-free water and 100% ethyl alcohol in a 10-mL reagent reservoir as shown in the following table, depending on the number of lanes used. Do not aliquot for single lane.

Component	1 lane (mL)	2 lanes (mL)	3 lanes (mL)	4 lanes (mL)	5 lanes (mL)	6 lanes (mL)	7 lanes (mL)	8 lanes (mL)
Nuclease-free water	0.38	2.00	2.00	2.50	2.50	3.00	3.50	4.00

Component	1 lane (mL)	2 lanes (mL)	3 lanes (mL)	4 lanes (mL)	5 lanes (mL)	6 lanes (mL)	7 lanes (mL)	8 lanes (mL)
100% ethyl alcohol	0.05	2.00	2.00	2.00	2.00	2.00	2.00	2.00

- 3 Load each lane used in the cartridge with the materials listed using the BD Rhapsody™ P8xP1200µL Pipette (or BD Rhapsody™ P1200µL Pipette).

Material to load	Volume (µL)	Pipette mode	Incubation at room temperature
Air	380	Prime/Wash	—
Nuclease-free water	380	Prime/Wash	1 min
Air	380	Prime/Wash	—
100% ethyl alcohol	50	EtOH Prime	—
Air	380	Prime/Wash	—

- 4 Use a lint-free wipe to remove liquid residue on the outside of the cartridge. Liquid residue inside the cartridge will not affect performance of the unused lanes.
- 5 Make sure the seals of the unused lanes are intact, place the cartridge for storage in the pouch provided with a desiccant bag, seal the double zipper bag, keep the cartridge flat, and store at room temperature in the dark.
- 6 Clean the BD Rhapsody™ HT Xpress System with 10% bleach or 70% ethyl alcohol.
- 7 Appropriately dispose of the waste collection container, unused cartridge buffers, and cartridge if all eight lanes have been used.



Partially used cartridges can be stored for up to 1 year at room temperature.

Troubleshooting

For additional troubleshooting on scanning or cartridge loading, see the troubleshooting section in the instrument user guide.

For technical support, contact your local Field Application Specialist (FAS) or scomix@bd.com.

Observation	Possible causes	Recommended solutions
Reported viability from the BD Rhapsody™ Scanner suspected to be too high.	DRAQ7™ staining in the current protocol is optimized for cell lines. The optimal concentration of DRAQ7™ might be higher.	Before the BD Rhapsody™ experiment, optimize the DRAQ7™ concentration for your cell types according to the manufacturer's protocol. See the DRAQ7™ technical data sheet, 564904.pdf .
Cannot move the RETRIEVAL slider.	The front slider is in the WASTE position.	Slide the front slider to the BEADS position.

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